

AquaRNA™ Instruction Manual

General Information

Description

AquaRNA™ (formerly AquaStool™, AquaPreserve™) is a multifunctional aqueous reagent for fecal sample stabilization, DNA and RNA extraction and PCR inhibitor removal. It may be used to extract fecal DNA for non-invasive genotyping of transgenic animals or preserve and extract DNA/RNA from human and animal stools for host and gut microbiome research and viral, bacterial, fungal, and parasitic detection. Additionally, AquaRNA may also be used to extract total DNA/RNA from various biospecimens, including cultured cells, microbes, blood, saliva, and animal or plant tissues.

Specification

Product Name	AquaRNA™ Solution
Product #	AR5030M
Size	30 ml AquaRNA™ for 200 mouse pellet extractions
MSDS	Available at www.boltii.com
Storage	Store tightly capped at 22 °C. Vortex to mix well before dispensing.
Note	Order these ancillary reagents with AquaRNA™ for specific applications: <ul style="list-style-type: none">• AquaRemove™ (AR1208M) for human fecal DNA/RNA• ProSink™ (RS9015M) for blood, liver, pancreas and spleen DNA/RNA

Terms & Condition

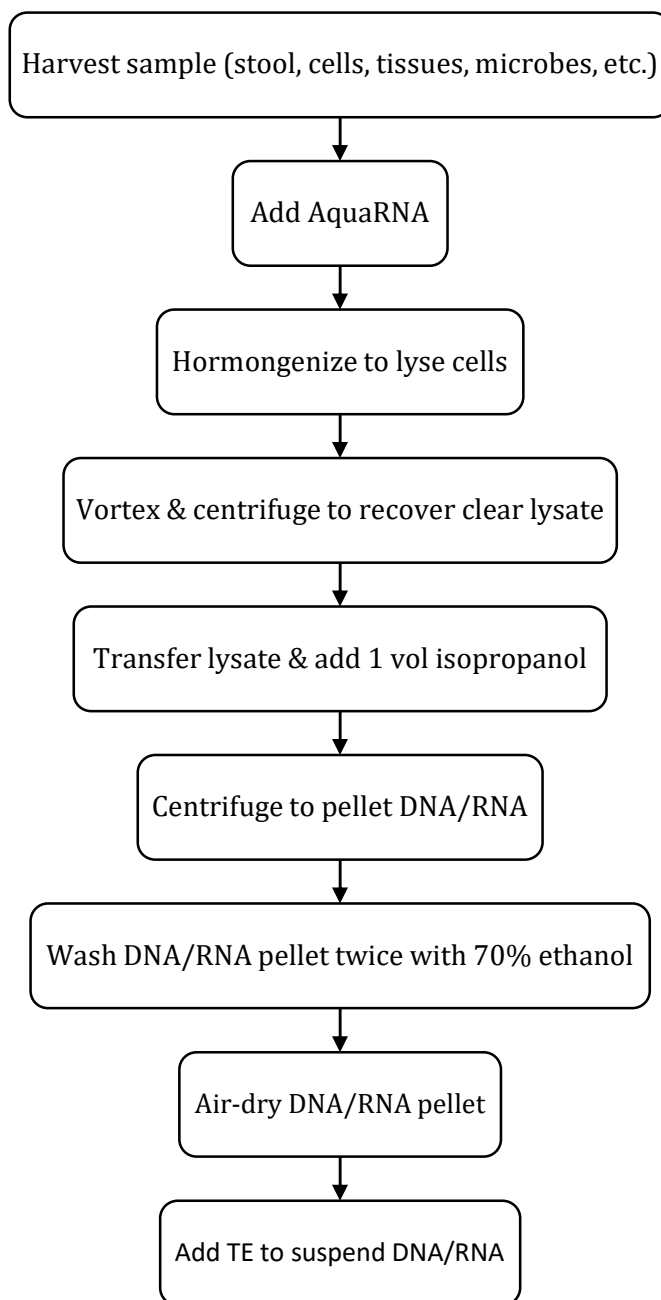
Product Usage: For *In Vitro* Laboratory Research Use Only. NOT to be administered to humans or used for medical diagnosis.

Safety, Handling, and Warnings: Contains guanidine thiocyanate, is harmful if swallowed and causes irritation to skin, eyes and respiratory tract. Do not mix with Bleach.

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AquaRNA™ Protocol Flowchart

Cultured Cell DNA/RNA Protocol

Extract DNA/RNA from cultured cells (~1–2 million cells).

1. Harvest Cells

- Pellet $1-2 \times 10^6$ cells at 3,000×g for 5 min.
- Carefully aspirate all medium.

2. Cell Lysis

- Add **100 µL AquaRNA™** to the cell pellet.
- Vortex vigorously for 1 min until no visible clumps remain.
- Incubate at 22 °C for 15 min to lyse cells.

3. Pellet Debris

- Centrifuge at 12,000×g for 5 min.
- Transfer the clear supernatant to a new tube without disturbing the pellet.

4. DNA/RNA Precipitation

- To pellet **mRNA**: Add a quarter volume of isopropanol.
- To pellet **total DNA/RNA**: Add an equal volume of isopropanol.
- Vortex 30 s.
- Centrifuge 12,000×g for 5 min to pellet mRNA or total DNA/RNA.
- Flip tube to discard supernatant.

5. Ethanol Wash

- Rinse the pellet with **70% ethanol** (fill the tube, including cap).
- Discard ethanol and repeat once.
- Tap tube on paper towel to remove residual ethanol.
- Air-dry pellet for 15 min.

6. DNA/RNA Solubilization

- Add **100 µL TE buffer**; pipette and vortex to disperse.
- Incubate at 22 °C for 15 min to solubilize DNA/RNA.
- Spin 5 min and transfer the DNA/RNA-containing supernatant to a new tube.
- Store at -20 °C.

Mouse Fecal DNA Protocol

Extract fecal DNA from mouse fecal pellets for genotyping.

1. Sample Collection

- Place individual mouse in a cage with clean lining.
- Collect pellets in labeled tubes. Air-dry at 37 °C for 24 h if long-term storage is desired.

2. Lysis

- Add ~25 mg white sand and **150 µL AquaRNA™** in a 1.5-mL tube.
- Add a mouse pellet.
- Incubate 22–65 °C for 15–30 min to soften the pellet.
- Vortex/bead-beat sample thoroughly.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

7. PCR genotyping

- Centrifuge the frozen DNA solution to pellet any insoluble (contains PCR inhibitor).
- Use 1 µL clear DNA supernatant per 25-µL PCR reaction.

Human Fecal DNA/RNA Protocol

Extract fecal DNA/RNA from stool samples of human and other large animals.

1. Sample Collection

- Add ~1 g stool in 10 mL AquaRNA™ in a 15-mL tube.
- Smash and disperse the stool with a spoon or rod.
- Close the tube and shake vigorously.
- Ship at ambient temperature for DNA; use cold shipping for RNA.
- Store at -80 °C.

2. Lysis

- Thaw the stool sample and shake vigorously.
- Transfer 0.5 mL stool sample to a 1.5-mL tube preloaded with Add ~100 mg white sand.
- Vortex/bead-beat sample thoroughly.
- For better PCR inhibitor removal: Add 250 µL AquaRemove™ (1:1 diluted with isopropanol), vortex and incubate 5–10 min.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

Tissue DNA/RNA Protocol

Extract DNA/RNA from animal or plant tissues.

1. Prepare Sample

- Cut ~2 mm tissue cubes (~25 mg).

2. Lysis

- Add **500 µL AquaRNA™**.
- Homogenize with a pestle-and-tube homogenizer or a multi-channel bead beater.
- After homogenization, add 2-3 drops of isopropanol to quench foaming.
- For plant tissue: Add 250 µL AquaRemove™ (1:1 diluted with isopropanol), vortex and incubate 5–10 min.

Proceed through Steps 4–6 as described in the Cultured Cell DNA/RNA Protocol.

Microbial DNA/RNA Protocol

Extract DNA/RNA from bacteria or yeast.

1. Harvest Cells

- Pellet 1 mL overnight culture at 12,000×g for 1 min.
- Aspirate supernatant.

2. Cell Lysis

- Pre-treat bacteria with **lysozyme**, or yeast with **lyticase**.
- Pellet cells and remove supernatant.
- Add **500 µL AquaRNA™**.
- Incubate **22 °C for 15 min**.
- Vortex vigorously for 1 min.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

Viral DNA or RNA Protocol

Extract DNA or RNA from DNA or RNA virus.

1. Harvest Virions

- Centrifuge 50 mL infected culture at 12,000×g for 1 min.
- Transfer supernatant to new tube.
- Add 1 volume 20% PEG8000 in 2.5M NaCl to 5 volume viral supernatant.
- Vortex and centrifuge at 12,000g for 10 min to pellet the virions.
- Aspirate to remove PEG supernatant as completely as possible.

2. Virion Lysis

- Add **500 µL AquaRNA™** and vortex to suspend the viral pellet.
- Incubate **on ice for 15 min.**
- Vortex vigorously for 1 min.
- Transfer viral lysate to 1.5-mL tube.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

Saliva DNA/RNA Protocol

Extract DNA/RNA from human saliva.

1. Collection

- Saliva: Collect **500 µL** after cheek rubbing.

2. Lysis

- Mix **500 µL sample + 500 µL AquaRNA™**.
- Incubate **15 min at 22 °C**.
- Vortex/bead-beat sample thoroughly.
- Add 250 µL AquaRemove™ (1:1 diluted with isopropanol) and vortex.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

Whole Blood DNA/RNA Protocol

Extract total DNA/RNA from whole blood.

1. Collection

- Collect anticoagulated blood using standard procedures.

2. Lysis

- Add **250 µL AquaRNA™** to 250 µL fresh or frozen blood (don't thaw blood without AquaRNA if you intend to recover blood RNA).
- Vortex to thaw blood and incubate **15 min at 22 °C**.
- Add **125 µL ProSink** to precipitate proteins.
- Vortex and incubate at 22 °C for >30 min.

Proceed through Steps 3–6 as described in the Cultured Cell DNA/RNA Protocol.

Plasma DNA/RNA Protocol

Extract cell-free plasma or serum DNA/RNA.

1. Collection

- Prepare the plasma (or serum) from fresh whole blood using standard methods.
- Transfer 2x 0.5 ml plasma to 2x 2-mL microfuge tubes.

2. Extraction

- Add **0.5 mL AquaRNA™** to each; vortex and incubate **15 min at 22 °C**.

Proceed through Steps 4–6 as described in the Cultured Cell DNA/RNA Protocol.

Frequently Asked Questions

Please read through these questions carefully. The answers provide additional helpful tips and useful information for the successful use of AquaRNA.

1. How should I store the AquaRNA solution?

AquaRNA may be stored at 22 °C for 12 months. If AquaRNA becomes precipitated when exposed to low temperature, you may incubate it at 37-50 °C for 15-20 min to resolubilize the reagent.

2. Why shouldn't I use Bleach to disinfect AquaRNA preserved fecal specimen?

AquaRNA contains guanidine thiocyanate. It may react with Bleach (sodium hypochlorite) and release toxic gases.

3. How should I air-dry the mouse fecal samples?

Air-dried mouse fecal samples can be stored long term at room temperature for future genotype verification. To air-dry mouse fecal pellets, incubate the fecal pellets in an opened microfuge tube on a dry heat bloc at 37 °C for 24 hours.

4. Why is my DNA/RNA solution showing a strong absorption below A260?

It is likely due to trace amount of guanidine salt contamination. If it interferes with your downstream applications, you may further purify the extracted DNA/RNA with a silica spin column (e.g., a plasmid miniprep column). Simply add an equal volume of 4 M GuHCl and 1M NaOAc (pH unadjusted, ~7.0) to your DNA/RNA solution and load it into the spin column, centrifuge to allow DNA/RNA binding to the silica membrane, wash the column with 0.6 ml 75% ethanol, and elute the DNA/RNA in 50 µL deionized water or TE buffer.

5. My mouse transgene was not amplified well, how may I improve it?

Try the following to improve mouse fecal DNA amplification: (a) after freezing the fecal DNA solution at -20 °C, re-centrifuge it to pellet and remove any insoluble, which may contain PCR inhibitors; (b) reduce the amount of fecal DNA used per PCR reaction (i.e., try using 0.5, 0.25, 0.1 and 0.01 µL extracted fecal DNA per PCR reaction); (c) increase PCR cycles up to 65; (d) add 1 mM DTT and 0.1 mg/mL BSA to the PCR reaction; (e) use a gel imager to visualize faint amplicon bands; (f) use AquaRemove™ with AquaRNA to purify fecal DNA (see "Human fecal DNA/RNA extraction" for details); and (g) further purify the fecal DNA with a silica spin column as outlined in #4 Question and Answer above.